

Chemistry 277, Spring 2019

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University of Maryland, College Park

General Chemistry and Energetics

Final Exam (100 points)

Your Name: -Key-

Your SID #: _____

Your Section # or time: _____

May 16, 2019

You have 50 minutes for this exam.

Explanations should be concise and clear. There is extra space on the last page if you need it.

You will not need a calculator for this exam. No other study aids or materials are permitted.

Generous partial credit will be given, *i.e.*, if you don't know, guess.

Useful Equations:

$$\sigma_Y^2 = \lim_{N \rightarrow \infty} \frac{1}{N} [\sum_i (Y_i - \bar{Y})^2]$$

$$\text{pH} = -\log([H^+])$$

$$\sigma_Y^2 = \left(\frac{\partial Y}{\partial u}\right)^2 \sigma_u^2 + \left(\frac{\partial Y}{\partial v}\right)^2 \sigma_v^2 + \dots$$

$$R = 0.08206 \text{ L} \cdot \text{atm/mole K}$$

$$T^2 = 4\pi^2 a^3 / GM$$

$$\ln K_{eq} = -\Delta H^\circ / (RT) + \Delta S^\circ / R$$

$$R = 8.314 \text{ J/mole K} = 1.987 \text{ cal/mole K} = N_A k_B$$

$$SEM = \frac{\sigma}{\sqrt{n}}$$

$$^\circ\text{C} = ^\circ\text{K} - 273.15$$

$$P(\dot{v})dv = Cv^2 \exp(-mv^2/2kT) \quad \ln k = (-E_a/RT) + \ln A$$

$$\text{pH} = \text{pK}_a + \log([A^-]/[HA])$$

$$K_p = K_c(RT)^{\Delta n}$$

$$K_w = [H^+][OH^-] = 10^{-14}$$

$$\text{Absorbance} = \epsilon c \ell$$

$$PV = nRT$$

$$N(E) = N_0 \omega \exp(-E/k_B T)$$

$$\text{pK}_a = -\log(K_a)$$

$$\text{pH(e.p.)} = \frac{1}{2} (\text{pK}_{a1} + \text{pK}_{a2})$$

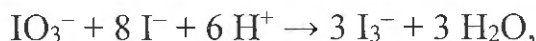
$$S = k_B \ln W$$

Honor Pledge: At the end of the examination time, please write out the following sentence and sign it, or talk to me about it:

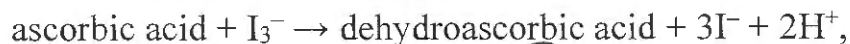
"I pledge on my honor that I have not given or received any unauthorized assistance on this examination."

1. Redox titration (25 pts)

In Lab 7B, we used iodate IO_3^- + excess iodide to generate triiodide according to:



then used the triiodide in excess to oxidize ascorbic acid, as in:



(a; 6 pts) Fill in the blanks:

...and then back-titrated with (word): (Mc) thiosulfate ⁽⁺³⁾ to find out how much triiodide was still left

according to this reaction: $\text{I}_3^- + 2 \text{S}_2\text{O}_3^{2-} \rightarrow 3 \text{I}^- + \text{S}_4\text{O}_6^{2-}$. ⁽⁺³⁾

(b; 4 pts) If you Google for ascorbic acid titration, you can find a few undergraduate labs that do an acid-base titration to determine the purity of commercial vitamin C tablets. Why is this inferior to the redox titration that we and most others use? ^{(+2) for idea of other ingredients}

Vitamin C tablets have excipients that are probably there for buffering to make it less acid for the mouth and stomach. Any such agents would interfere with an acid/base titration. Using the redox properties of AA is more specific. ^{(+2) for idea of specificity of redox}

(b; 3 pts) Why is ~~iodate~~ iodate used as the primary concentration standard in this lab, as opposed to triiodide or thiosulfate?

Iodate is a stable, presumably pure solid - the triiodide and thiosulfate are not stable upon storage. ^{(+3) for either/both}

(d; 3 pts) What result would you have obtained if the supplier had a QC problem and supplied KIO_3 at 90% purity, with the remainder being entirely soluble and inert? Circle the best answer:

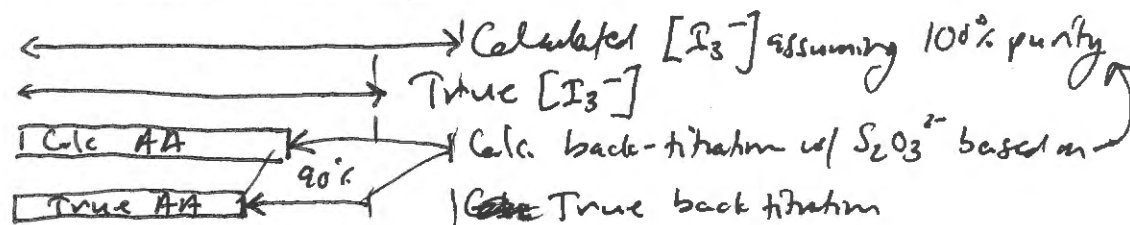
(i) There would have been no change in the experiment.

(ii) The calculated ascorbic acid concentration would be 10 % too low.

⁽⁺³⁾ (iii) The calculated ascorbic acid concentration would be 10 % too high.

(iv) The experiment would have failed completely.

(e; 3 pts) Briefly explain your answer to (d).



The true graph is just 90% of calculated in every (horizontal) line.

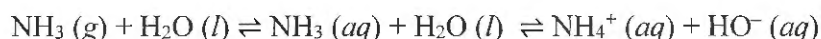
Calc. $[\text{AA}] \propto [\text{IO}_3^-]$ ^{(+3) for idea consistent w/ (d)}

(f; 6 pts) Briefly discuss how/why a truly terrible KIO_3 purity of ~15 % would have made (iv) the correct answer to part (d), and state what you could have done to recover.

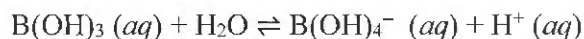
- +3 - If the I_3^- is not in excess there is nothing to back-titrate!
- +3 ~~each~~ for either one - Use a much smaller portion of the Vitamin C tablet and then ~~some~~ multiply to get the [AA]
- Will need some other primary standard to measure $[\text{KIO}_3]$.

2. Acid-base and methods development (16 pts)

Ammonia gas (NH_3) dissolves in water and is in equilibrium with ammonium hydroxide, but the pK_b of NH_3 is about 4.75 so the concentration of ammonium cation is low:



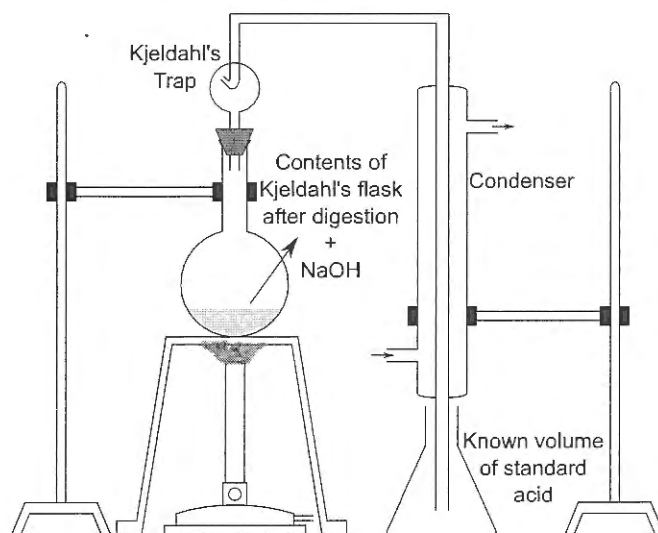
In the Kjeldahl assay for determining protein concentration, a sample is reacted completely with strong acid to convert all organic nitrogen to $\text{NH}_4^+ (\text{aq})$. The solution is then made basic, and whatever is volatile (i.e. not ions) is distilled into a receiving flask containing an acid, often the weak acid boric acid $\text{B}(\text{OH})_3$.



Then the ammonia and/or ammonium content in the receiving flask is measured.

(Not fun at all fact: The Kjeldahl assay can be fooled by the nitrogen-rich but non-nutritious and poisonous compound melamine, which was used in 2007-2008 by unscrupulous pet food and baby formula manufacturers to simulate protein in their products, killing dogs and babies.)

(a; 4 pts) Why is it necessary to raise the pH before doing the distillation?



Picture credit: By Roshan220195 - Own work, CC BY-SA 3.0, <https://commons.wikimedia.org/w/index.php?curid=18937792>

The $\text{NH}_3/\text{NH}_4^+$ must be in the volatile

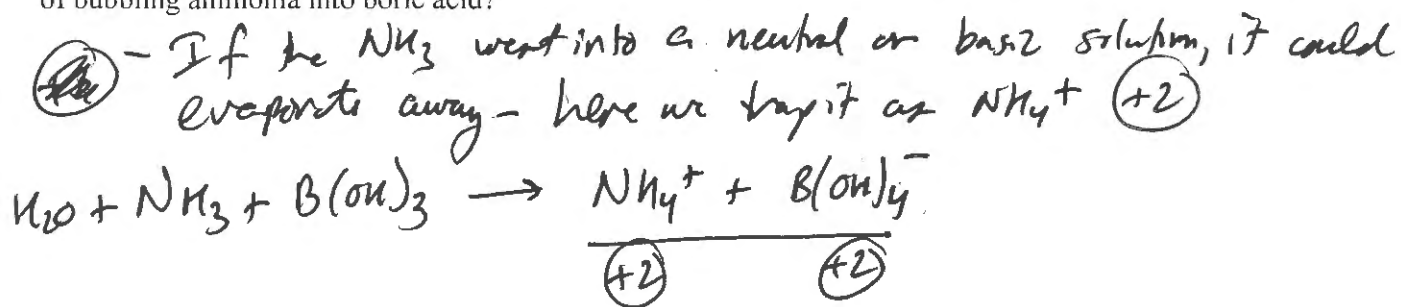
NH_3 form or it will not distill over \rightarrow go to high pH to deprotonate all NH_4^+ .

+2 for idea of volatility

+2 for idea of NH_3 @ high pH, implicit OK

Score for the page 10

(b; 6 pts) Why is it necessary to collect the distillate in an acidic receiving solution? What are the products of bubbling ammonia into boric acid?



(c; 6 pts) Experimentally, how could you determine the amount of ammonia that ended up in the collection flask?

- There is some mixture of NH_4^+ and $\text{B}(\text{OH})_4^-$ and $\text{B}(\text{OH})_3$ in the mix. flask.

(+4) for either - Titrate w/ HCl or other acid (like $\text{B}(\text{OH})_3$...) to convert all $\text{B}(\text{OH})_4^-$ back to $\text{B}(\text{OH})_3 \rightarrow$ that is the amount of NH_4^+

- or titrate w/ NaOH to measure amt of $\text{NH}_4^+ + \text{B}(\text{OH})_3$

(+2) for recognizing this issue - Another way you need to know the initial $[\text{B}(\text{OH})_3]$ or be prepared to disentangle 2 equivalence points

3. SDL (12 pts)

Describe an unexpected finding from your SDL, why it was a surprise or disappointment, and how you would either make the issue go away by improving your methods or else investigate it further.

(+4) for the finding "NPs were grey instead of colored"

(+4) for why the surprise "wanted beautiful color as in the literature"

(+4) for "Do TBuM to see if the particles are polydispersed"

next "Reflex the way the prof said"

step. "Try a different capping agent"

4. Nanoparticles and Chelators (22 pts)

(a; 6 pts) Write the name of the physical phenomenon that causes nanoparticles of different sizes to be

different colors: Surface plasmon resonance (+3). What technique provided the calibration curve we used to estimate size from λ_{\max} ? transmission electron microscopy (+3) (TEM +2)(b; 3 pts) What color do you get if all wavelengths are absorbed equally? gray (or black ok) (+3)In Lab 6, we used EDTA to rip Zn^{+2} away from a xylenol orange indicator.

(c; 4 pts) Explain why we used EDTA in this lab instead of just titrating directly with xylenol orange until no further color change was observed.

(+2) for any reason for { It is hard to tell the difference between more of a yellow color and a change to an orange color - the end point would not be obvious. (+2)

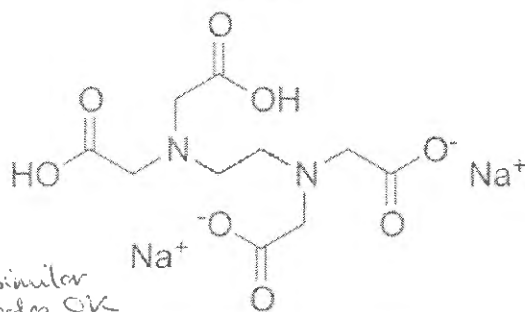
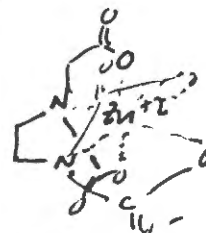


Figure 1: The Chemical Structure of Disodium EDTA

<https://pediaa.com/difference-between-disodium-edta-and-tetrasodium-edta/>(d; 3 pts) Explain why EDTA has such a strong binding affinity for Zn^{+2} (and many other metal ions).

Chelation - it can make multiple non-covalent interactions with the ligand. (+3)
or +3 for more words



or +3 for just a picture that gets the idea across

(e; 6 pts) Explain why EDTA is used in the treatment of lead and mercury poisoning, but why large doses should be avoided unless one has actually been poisoned.

(+3) - EDTA can chelate Pb^{+2} or Hg^{+2} so it can be removed from the body.

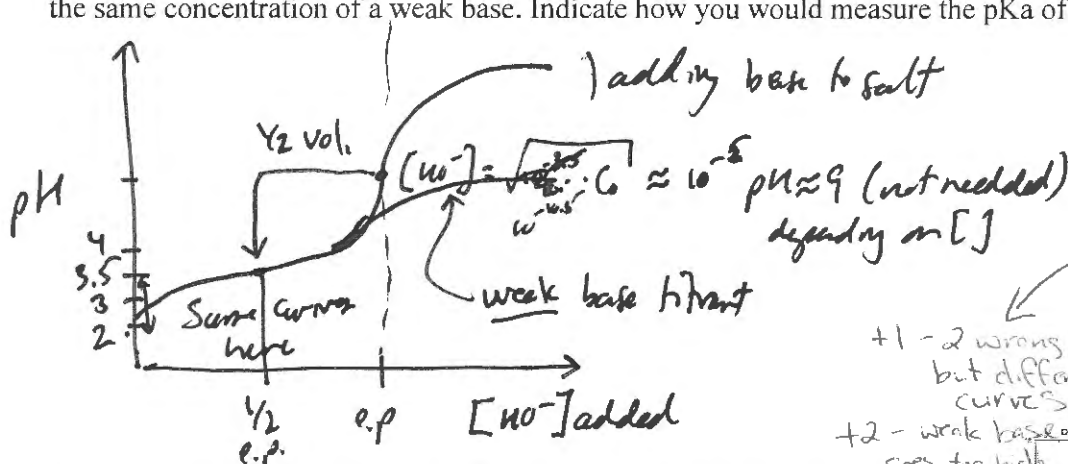
(+3) - But it can also remove useful metal ions like Ca^{+2} and Mg^{+2} , so taking too much can be dangerous.

Score for the page

1/22
22

5. Using the computer as if it were a piece of apparatus (25 pts)

(a; 12 pts) Sketch a curve showing pH vs. volume of base added with two titration curves, one curve for an acid with a pK_a of 3.5 with strong base, and the other curve for the same acid at the same concentration with the same concentration of a weak base. Indicate how you would measure the pK_a of the acid.



(+3) for idea of titration
(+2) for $pH = pK_a$ at $1/2$ e.p.

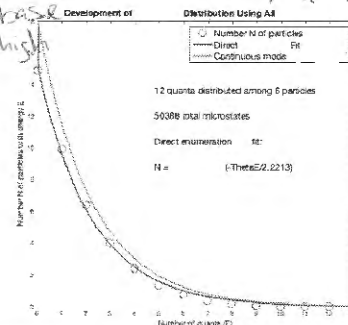
(+3) for 2nd curve correct

(+3) for indicating to log $1/2$ the volume at the e.p.
→ +2 for say $1/2$ e.g. pH

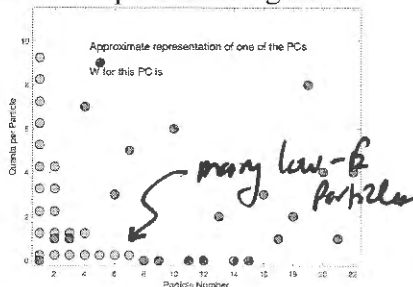
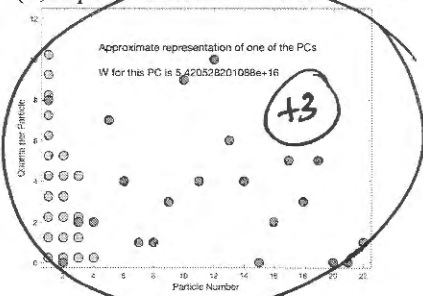
+1 - 2 wrong but different curves
+2 - weak base goes too high

(b; 4 pts) What is the functional form of the curve that is fit to the observed number of particles that have a given number of quanta in the plot shown at the right, and whose distribution is shown?

(decaying) exponential Boltzmann
(+2) → +1 for compare ideal to experimental
(+2)



(c; 3 pts) Circle the distribution below that represents a higher T :



(d; 6 pts) How would you exercise the `micro_mvcr3` program to see whether its toy model for entropy is an extensive state function (which is the idea that two moles of a pure substance at a given temperature has twice the entropy of one mole)?

- Run the program w/ e.g. $g = 30$ and $p = 20$, record the entropy
(+3) for idea

- Run the " w/ e.g. $g = 60$ and $p = 40$ - same g/p ratio i.e. just 2x the same system → does the entropy double or not?

(+3) for looking at results for S

$\frac{S}{G}$ if $\uparrow p$ and g but not mention double

Page	Score
2	/19
3	/10
4	/24
5	/22
6	/25
Total	/100

Score for the page

/25

CELLO

9

37

43

48

153

158

163

169

cresc.

f

dim.

sempre dim.

p

sempre

pp

pizz.

p

G

II

Detailed description of the musical score: The score is for a cello part. It begins on staff 37 with a treble clef and a key signature of one sharp (F#). The time signature is 3/4. The music consists of eighth and sixteenth notes, often beamed together. Staff 43 shows a circled 'F' and a 'cresc.' marking. Staff 48 has a circled 'G' and a 'f' marking. Staff 153 has a 'dim.' marking. Staff 158 has a 'sempre dim.' marking. Staff 163 has a 'p' marking, a circled 'G', and a 'sempre' marking. Staff 169 has a 'pizz.' marking and a 'p' marking. A bracketed section on staff 163 is marked 'II'.